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Review Article

RECENT POTENTIAL USAGE OF SURFACTANT FROM MICROBIAL ORIGIN IN PHARMACEUTICAL AND BIOMEDICAL ARENA: A PERSPECTIVE

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ABSTRACT

The use and potential commercial application of biosurfactant has increased during the past decade which can be used as emulsifiers, deemulsifiers, wetting and foaming agents, functional food ingredients and as detergents in petroleum, petrochemicals, environmental management, agrochemicals, foods and beverages, cosmetics and pharmaceuticals and in the mining and metallurgical industries. Their antibacterial, antifungal and antiviral activities make them relevant molecules for applications in combating many diseases and as therapeutic agents. In addition to this their role as antiadhesive agents against several disease causing pathogens makes their utility as suitable antiadhesive coating agents for medical insertional materials which helps in the reduction in a large number of hospital infections without the use of synthetic drugs and chemicals. This review looks at the various pharmaceutical, biomedical and therapeutic perspectives on biosurfactant applications.

Key words: Biosurfactant, Antiadhesive agent, Antimicrobial activities, Antiviral activities,

INTRODUCTION

Biosurfactants (Microbial Surface Active Agents) have become recently an important product of biotechnology industrial. pharmaceutical and biomedical applications. These are the amphiphilic compounds produced mostly on microbial cell surfaces or may be excreted extracellularly. This contains both hydrophobic and hydrophilic groups that confer the ability to accumulate between fluid phases and thus it is used in reduction of interfacial tension¹. Unlike chemical surfactants, which are mostly derived from petroleum feedstock, these molecules can be produced by microbial fermentation processes using cheaper agro-based substrates and waste materials. During the past few years, biosurfactant production various microorganisms has been studied extensively². They are biodegradable and less toxic and are effective at extreme temperatures and pH values³.

Most of the work on biosurfactant applications has been focusing on bioremediation of pollutants as it enhances the solubility and availability of hydrophobic pollutants, thus increasing their potential for biodegradation and also applied in the process of microbial enhanced oil recovery ^{4,5}. Although the production of biosurfactants has steadily increased during the past decade the large scale production of these molecules has not been realized satisfactorily because of its high recovery and purification costs⁶. Their antibacterial, antifungal and

antiviral activities make them as a relevant molecules for applications in combating many diseases and as a good therapeutic agent in recent days, so its draws the attention in the pharmaceutical and biomedical field⁷. Moreover they can be used as anti adhesive agents to the diseasing causing pathogens⁸.

Here, we discuss some of the new and exciting applications and related developments of various microbial surfactants in the field of pharmaceutical and biomedical sciences and also to provide an overview of biosurfactant activities that could be exploited further in developing alternative drugs in the coming future.

Biosurfactant Classification

Biosurfactants are generally classified mainly by their chemical composition and microbial origin. It can be divided into low molecular mass molecules (providing lower surface and interfacial tension) and high molecular mass polymers (can be used as emulsion stabilizing agents)⁹. The brief classification of biosurfactant with their microbial sources has been summarized in the table-1.

Various Biological Activity of Biosurfactant

Biosurfactant have so many potential biological activities like anti cancer activity, anti microbial activity, anti adhesiveness, micro bubbles stabilization activity, anti-HIV and sperm immobilizing activity, immunomodulatory and various therapeutic activities which has been listed in the table-2.

Anti-cancer activity

activity of The biological several groups biosurfactants like glycolipids. including mannosylerythritol lipids-A, mannosylerythritol lipids-B, polyol lipid, rhamnolipid, sophorose lipid, succinoyl trehalose lipid-1 and succinoyl trehalose lipid-3 have been investigated¹⁹ .Except rhamnolipid, all these groups biosurfactants were found to induce differentiation instead of cell proliferation in the human pro-myelocytic leukaemia cell line²⁰. erythritol lipid is a group of surfactant glycolipid derived from Candida antartica enhances the differentiation of granulocytes. This glycolipid also induces the stage of apoptosis and differentiation of mouse malignant melanoma cells. The cytotoxic effects of sophorolipids on cancer cells of H7402, A549, HL60 and K562 were investigated by MTT assay in which a dose dependent inhibition ratio was noticed on cell viability ^{21, 22}.

Anti-microbial activity

The antimicrobial activity of two biosurfactants obtained from probiotic bacteria namely Lactococcus lactis-53 and Streptococcus thermophilus-A, against a variety of bacterial and yeast strains²³. The lipopeptide iturin from Bacillus subtilis showed potent antifungal activity²⁴. A rhamnolipid mixture obtained from *Pseudomonas* aeruginosa-AT10 showed inhibitory activity against so many microorganisms like Escherichia coli. Micrococcus luteus. Alcaligenes faecalis, Mycobacterium phlei. It also possesses the antifungal properties against Aspergillus niger, Chaetonium globosum and the strain of Aureobasidium pullulans 25. The mannosyl erythritol lipid is a group of glycolipid obtained from Candida antartica has demonstrated antimicrobial activity particularly against Gram-positive bacteria²⁶.

Biosurfactant as Anti-adhesives

The probiotic strains like *Lactobacillus plantarum*-299v and Lactobacillus rhamnosus-GG inhibit the adhesion of Escherichia coli to the intestinal epithelial cells by stimulating epithelial expression of mucins as these are the key producer of biosurfactant ²⁷. A surfactant produced by Streptococcus thermophilus has been used for fouling control of heat-exchanger plates in pasteurizers, as it can be able to retard the colonization of other thermophilic strains of Streptococcus which are responsible for fouling. More over pretreatment of stainless steel surfaces with a biosurfactant obtained from Pseudomonas fluorescens inhibits the colonization and adhesion of Lactobacillus monocytogene- L028 strain²⁸. It was observed that *Rothia dentocariosa*, which is one of the strains responsible for the failure of valve prosthesis but this can be avoided when this prosthesis is

exposed to biosurfactant obtained from Streptococcus thermophilus-A. Recently it was also demonstrated that the rate of microbial adhesion with a rhamnolipid biosurfactant containing solution was significantly reduced for a variety of bacterial and yeast strains isolated from explanted voice prostheses to silicone rubber²⁹. A biosurfactant of *Pseudomonas fluorescens* was found to inhibit the adhesion of Listeria monocytogenes-LO28 to the stainless surfaces.Lactobacilli sps. important are in the maintenance of the healthy urogenital flora. There are some reports of inhibition of biofilm formed by uropathogens and yeast on silicone rubber by biosurfactants produced by Lactobacillus acidophilus 33,

Biosurfactant for the Stabilization of Micro bubbles

Currently, applications of biosurfactant-based micro bubbles are focused mainly on ultrasound diagnosis and therapy³⁰. Extensive studies have been performed on the use of micro bubbles as ultrasound contrasting agents. These types of micro bubbles are usually encapsulated in a shell of surfactant, protein, lipid or polymer to increase stability with *in vivo* condition. Their advantages include fragility when exposed to moderate energy ultrasound and ease of preparation. A wide range of substances such as drugs, DNA and virus particles can be bound to the shells of the micro bubbles with biosurfactant making them potential delivery systems for drugs and genes^{31,32}.

Anti-HIV and Sperm Immobilizing activity of Biosurfactant

The increased occurrence of human immunodeficiency virus (HIV) in women aged groups of 15–49 years have been identified as the urgent demand for a female-controlled effective and safe vaginal topical microbicide. The sophorolipid produced by *Candida bombicola*³³ and its structural analogues have been studied and proved its spermicidal, anti-HIV and cytotoxic activities. The sophorolipid diacetate ethyl ester derivative is the most potent spermicidal and virucidal agent of the series of sophorolipids studied³⁴. Its virucidal activity against HIV and spermimmobilizing activity against human semen are similar and equivalent to those of nonoxynol-9³⁵.

Sterilizing agents in surgical

The biosurfactant from *Lactobacillus fermentum* was reported to inhibit *Staphylococcus aureus* infection and its adherence to surgical implants. More over Surfactin which is obtained from *Streptococcus thermophilus* able to decrease the amount of biofilm formation by *Salmonella typhimurium* and *Proteus mirabilis* in PVC ³⁶. Pre-treatment of silicone rubber with *Streptococcus*

thermophilus surfactant can able to inhibit the adhesion of Candida albicans-87 up to 85%, whereas surfactants from Lactobacillus fermentum and Lactobacillus acidophilus adsorbed on glass surface can inhibit the number of adhering uropathogenic cells of Enterococcus faecalis up to 77% ³⁷.

Immuno modulatory action of Biosurfactant

Sophorolipids are the best modulators in the immune response. It has been demonstrated previously that sophorolipids decreased sepsis related mortality at the interval of 36 hours *in vivo* in a rat model of septic peritonitis by adhesion molecules and cytokine production and also decreasing the IgE production *in vitro* in U266 cells by affecting plasma cell activity. The results show that sophorolipids is down regulating important genes involved in IgE-pathobiology in order to decrease IgE production in U266 cells. Hence the sophorolipids can be used as an anti-inflammatory agent and can be a novel potential therapeutic in the process of altered IgE regulation³⁸.

Therapeutic Applications of Biosurfactant

In order to generate monoclonal antibodies for the serological detection of drugs, antibodies and toxins a suitable and effective adjuvant is needed. It was proved that bacterial lipopeptides constitute potent nontoxic and nonpyrogenic immunological adjuvant when mixed with conventional antigens. A group of synthetic lipopeptide (N-palmitoyl-S-[2, 3-bis (palmitoyloxy)-(2R, S)-propyl-]-(R)-cysteinyl-serine) which is coupled to a T_h cell epitope efficiently enhanced the specific immune response against low molecular weight compounds in different species. A marked enhancement of the humoral immune response was obtained with the low molecular mass antigens with iturin, herbicolin and microcystin which are coupled to poly-L-lysine in rabbits and in chickens³⁹.One of the most important groups of organisms being studied for their use as an effective probiotic are members of the Lactobacilli group, which have the potential to prevent pathogen colonization and help to restore the normal microbial flora⁴⁰.

In spite of the immense potential of the biosurfactants in this field, their use still remains limited, possibly because of their comparatively high production cost, as well as their toxicity towards human systems. More research on human cells and human natural micro biota required to be performed in order to justify the potential need of biosurfactants in the biomedical area.

CONCLUSION

A host of interesting features of biosurfactants have led to a wide range of potential applications in the pharmaceutical and biomedical field. They are useful as antibacterial, antifungal and antiviral agents, and they

have the potential for immunomodulatory molecules adhesive agents and in vaccines and gene therapy. Successful commercialization of every biotechnological product mainly depends largely upon its bioprocess economics. At present, the prices of microbial surfactants are not competitive with those of the chemical surfactants due to their high production costs and low yields. Hence, they have not been commercialized extensively. For the production of commercially viable biosurfactants, process optimization at the biological and engineering level needs to be improved. Improvement in the production technology of biosurfactants has already enabled a 10 to 20 fold increase in productivity, although further significant improvements are required.

However, the use of cheaper substrates for the production of biosurfactant with recombinant and mutant hyper producing microbial strains and optimal growth and production conditions having efficient multi-step downstream processing methods can make biosurfactant production economically feasible in the coming future.

REFERENCES

- 1. Karanth NGK, Deo PG, Veenanadig NK. Microbial production of biosurfactants and their importance. Current Science, 1999, 77: 116–123.
- 2. Banat IM, Makkar RS, Cameotra SS. Potential commercial applications of microbial surfactants. Applied Microbiology and Biotechnology, 2000, 53: 495–508.
- 3. Cameotra SS, Makkar RS. Synthesis of biosurfactants in extreme conditions. Applied Microbiology and Biotechnology,1998, 50: 520-529.
- 4. Mulligan CN. Environmental applications for biosurfactants. Environmental Pollution, 2005, 133:183–198.
- 5. Banat IM. Biosurfactants production and possible uses in microbial enhanced oil recovery and oil pollution remediation: A review. Bioresource Technology, 1995, 51: 1–12.
- 6. Maneerat S. Production of biosurfactants using substrates from renewable-resources. Songklanakarin journal of Science and Technology, 2005, 27: 675–683.
- 7. Ron EZ, Rosenberg E. Natural roles of biosurfactants. Environmental Microbiology,2001,3: 229–236.
- 8. Pooja S, Swaranjit SC. Potential applications of microbial surfactants in biomedical sciences. TRENDS in Biotechnology , 2004, 22: 142-145.
- 9. Rosenberg E, Ron EZ. High- and low-molecular-mass microbial surfactants. Applied Microbiology and Biotechnology, 1999, 52: 154-162
- 10. Poremba K, Gunkel W, Lang S, Wagner F. Toxicity testing of synthetic and biogenic surfactants on marinemicroorganisms. Environmental Toxicology and Water Quality, 1991, 6:157–163.
- 11. Schulz D, Passeri A, Schmidt M, Lang S, Wagner F, Wray V, et al. Marine biosurfactants. I. Screening for biosurfactants among crude oil degrading marine microorganisms from the North Sea. Zeitschrift für Naturforschung C, 1991, 46:197–203.
- 12. Passeri A, Lang S, Wagner F, Wray V. Marine biosurfactants. II: Production and characterization of an anionic trehalose tetraester from the marine bacterium *Arthrobacter sp.* EK1. Zeitschrift für Naturforschung C, 1991, 46:204–209.

- 13. Das P, Mukherjee S, Sen R. Antimicrobial potential of a lipopeptide biosurfactant derived from a marine *Bacillus circulans*. Journal of Applied Microbiology, 2008, 04:675–684.
- Mukherjee S, Das P, Sivapathasekaran C, Sen R. Antimicrobial biosurfactants from marine *Bacillus circulans*: extracellular synthesis and purification. Letters in Applied Microbiology, 2009, 48: 281–288.
- Yakimov MM, Timmis KN, Wray V, Fredrickson HL.
 Characterization of a new lipopeptide surfactant produced by thermotolerant and halotolerant subsurface *Bacillus licheniformis*.
 BAS50. Applied Environmental Microbiology, 1995, 61:1706– 1713.
- 16. Maneerat S, Nitoda T, Kanzaki H, Kawai F. Bile acids are new products of a marine bacterium, Myroides sp. strain SM1. Applied Microbiology and Biotechnology, 2005,67:679–683.
- 17. Peng F, Liu Z, Wang L, Shao Z. An oil-degrading bacterium: *Rhodococcus erythropolis* strain 3C-9 and its biosurfactants. Journal of Applied Microbiology, 2007, 102:1603–1611.
- 18. Pepi M, Cesaro A, Liut G, Baldi F. An Antarctic psychrotrophic bacterium *Halomonas sp.* ANT-3b, growing on n-hexadecane, produces a new emulsifying glycolipid. FEMS Microbiology Ecology, 2005, 53:157–166.
- 19. Isoda H, Shinmoto H, Matsumura M, Nakahara T. The neurite initiating effect of microbial extracellular glycolipids in PC12 cells. Cytotechnology, 1999, 31:163–170.
- 20.Zhao X, Wakamatsu Y, Shibahara M, Nomura N, Geltinger C, Nakahara T et al. Mannosylerythritol lipid is a potent inducer of apoptosis and differentiation of mouse melanoma cells in culture. Cancer Research, 1999, 59: 482–486.
- 21. Wakamatsu Y, Zhao X, Jin C, Day N, Shibahara M, Nomura N, et al., Mannosylerythritol lipid induces characteristics of neuronal differentiation in PC12 cells through an ERK related signal cascade. European Journal of Biochemistry, 2001, 268: 374–383.
- 22. Chen J, Song X, Zhang H, Qu Y. Production, structure elucidation and anticancer properties of sophorolipid from *Wickerhamiella domercqiae*. Enzyme and Microbial Technology, 2006, 39: 501–506.
- 23. Rodrigues LR, van der Mei HC, Teixeira J. Influence of biosurfactants from probiotic bacteria on formation of biofilms on voice prostheses. Applied Environmental Microbiology, 2004, 70: 4408–4410.
- 24. Besson F, Peypoux F, Michel G,Delcambe L. Characterization of iturin A in antibiotics from various strains of *Bacillus subtilis*. Journal of Antibiotics, 1976, 29:1043–1049.
- 25. Abalos A, Pinazo A, Infante MR, Casals M, Garcýa F, Manresa A. Physicochemical and antimicrobial properties of new rhamnolipids produced by *Pseudomonas aeruginosa* AT10 from soybean oil refinery wastes. Langmuir, 2001, 17:1367–1371.
- 26. Kitamoto D, Yanagishita H, Shinbo T, Nakane T, Kamisawa C, Nakahara T. Surface active properties and antimicrobial activities of mannosylerythritol lipids as biosurfactants produced by *Candida antarctica*. Journal of Biotechnology, 1993, 29: 91–96.
- 27.Mack DR, Michail S, Wei S, McDougall L, Hollingsworth MA. Probiotics inhibit enteropathogenic *E. coli* adherence in vitro by inducing intestinal mucin gene expression. The American Journal of Physiology, 1999,276: 941–950.
- 28. Hood SK, Zottola EA. Biofilms in food processing. Food Control, 1995, 6:9-18.
- 29. Rodrigues LR, Banat IM, Van der Mei HC. Interference in adhesion of bacteria and yeasts isolated from explanted voice prostheses to silicone rubber by rhammolipid biosurfactants. Journal of Applied Microbiology, 2006, 100:470-480.

- 30. Wright JR. Pulmonary surfactant: a front line of lung host defense. Journal of Clinical Investigation, 2003, 111: 1453–1455.
- 31. Morawski AM, Lanza GA, Wickline SA. Targeted contrast agents for magnetic resonance imaging and ultrasound. Current Opinion in Biotechnology, 2005, 16: 89–92.
- 32. Bloch SH, Short RE, Ferrara KW, Wisner E R. The effect of size on the acoustic response of polymer-shelled contrast agents. Ultrasound in Medical and Biology, 2005, 31: 439–444.
- 33. Meylheuc T, Van Oss CJ, Bellon-Fontaine MN.Adsorption of biosurfactant on solid surfaces and consequences regarding the bioadhesion of *Listeria monocytogenes* LO28. Journal of Applied Microbiology, 2001, 91:822–832.
- 34. Velraeds MM, van de Belt-Gritter B, van der Mei HC, Reid G, Busscher HJ. Interference in initial adhesion of uropathogenic bacteria and yeasts to silicone rubber by a *Lactobacillus acidophilus* biosurfactant. Journal of Medical Microbiology,1998, 47: 1081–1085.
- 35. Shah V, Doncel GF, Seyom T, Eaton KM, Zalenskya I, Hagver R, Gross R. Sophorolipids, microbial glycolipids with anti-human immunodefficiency virus and sperm-immobilising activities. Antimicrobial Agents and Chemotherapy, 2005, 49: 4093–4100.
- 36. Gan BS, Kim J, Reid G, Cadieux P, Howard JC. *Lactobacillus fermentum* RC-14 inhibits *Staphylococcus aureus* infection of surgical implants in rats. Journal of infectious Disease, 2002, 185:1369–1372.
- 37. Mireles JP, Toguchi A, Harshey RM. Salmonella enteric serovar typhimurium swarming mutants with altered biofilmforming abilities: Surfactin inhibits biofilm formation. Journal of Bacteriology, 2001, 183:5848–5854.
- 38. Hagler M, Smith-Norowitz TA,Chice S,Wallner SR,Viterbo D, Mueller CM et al. Sophorolipids decrease IgE production in U266 cells by down regulation of BSAP (Pax5), TLR-2, STAT3 and IL-6. Journal of Allergy Clinical Immunology, 2006, 119: 245–249.
- 39. Mittenbühler K, Loleit M, Baier W, Fischer B, Sedelmeier E, Jung G, et al.Drug specific antibodies: T-cell epitopelipopeptide conjugates are potent adjuvants for small antigens *in vivo* and *in vitro*. International Journal of Immunopharmacology, 1997, 19:277–287.
- 40. Lang S, Wullbrandt D. Rhamnose lipids biosynthesis, microbial production and application potential. Applied Microbiology and Biotechnology, 1999, 51: 22–32.
- 41. Maier R, Soberon-Chavez G. *Pseudomonas aeruginosa* rhamnolipids: biosynthesis and potential applications. Applied Microbiology and Biotechnology, 2000, 54: 625–633.
- 42. Kameda Y, Ouchira S, Matsui K. Antitumor activity of *Bacillus natto* V. Isolation and characterization of surfactin in the culture medium of *Bacillus natto* KMD 2311. Chemical and Pharmaceutical Bulletin,1974,22: 938–944.
- 43. Bernheimer A, Avigad L. Nature and properties of a cytolytic agent produced by *Bacillus subtilis*. Journal of General Microbiology,1970, 61: 361-369.
- 44. Sheppard JD, Jumarie C, Cooper DG. Ionic channels induced by surfactin in plannar lipid bilayer membranes. Biochimica et Biophysica Acta ,1991,1064: 13–23.
- 45. Itokawa H, Miyashita T, Morita H. Structural and conformational studies of [Ile7] and [Leu7] surfactins from *Bacillus subtilis*. Chemical and Pharmaceutical Bulletin,1994,42: 604–607.
- 46. Mittenbuhler K, Loleit M, Baier W.Drug specific antibodies: T-cell epitope-lipopeptide conjugates are potent adjuvants for small antigens *in vivo* and *in vitro*. International Journal of Immunopharmacology, 1997,19: 277–287.
- 47. Naruse N, Tenmyo O, Kobaru S et al. Pumilacidin, a complex of new antiviral antibiotics: production, isolation, chemical properties,

- structure and biological activity. Journal of Antibiotics, 1990, 43: 267-280.
- 48. Shibahara M, Zhao X, Wakamatsu Y. Mannosylerythritol lipid increases levels of galactoceramide in and neurite outgrowth from PC12 pheochromocytoma cells. Cytotechnology, 2000, 33: 247–251.
- 49 . Zhao X, Geltinger C, Kishikawa S . Tretament of mouse melanoma cells with phorbol 12-myristate 13-acetate counteracts mannosylerythritol lipid-induced growth arrest and apoptosis. Cytotechnology, 2000, 33: 123–130.
- 50. Zhao X, Wakamatsu Y, ShibaharaM . Mannosylerythritol lipid is a potent inducer of apoptosis and differentiation of mouse melanoma cells in culture. Cancer Research, 1999,59: 482–486.
- 51. Busscher HJ, Van Hoogmoed CG, Geertsema-Doornbusch GI. Streptococcus thermophilus and its biosurfactants inhibit adhesion

- by Candida spp. on silicone rubber. Applied Environmental Microbiology, 1997, 63: 3810–3817.
- 52. Busscher HJ, Van de Belt-Gritter B, Westerhof M. Microbial interference in the colonization of silicone rubber implant surfaces in the oropharynx: *Streptococcus thermophilus* against a mixed fungal/bacterial biofilm. In: Rosenberg E, ed. Microbial Ecology and Infectious Disease. Washington, DC: American Society for Microbiology, 1999, 66–74.
- 53. Yakimov M, Timmis K, Wray V. Characterization of a new lipopeptide surfactant produced by thermotolerant and halotolerant subsurface *Bacillus licheniformis* BAS50. Applied Environmental Microbiology, 1995, 61: 1706–1713.
- 54. Grangemard I, Wallach J, Maget-Dana R . Lichenysin: a more efficient cation chelator than surfactin. Applied Biochemistry and Biotechnology, 2001, 90: 199–210

Table-1: Classification of different microbial surfactants.

Biosurfactant Classes	Sub Classes	Source	References
Glycolipids	Rhamnolipids	Pseudomonas aeruginosa	10
	Trehalose lipids	Rhodococcus erithropolis	
	Mannosylerythritol lipids	Candida antartica	10
			12
Lipopeptides	Surfactin/iturin	Bacillus subtilis	13
	Lichenysin	B. licheniformis	
	Serrawettin	Serratia marcescens	14
			15
Phospholipids	Corynebacterium lepus		16
Surface-active antibiotics	Gramicidin	Brevibacterium brevis	11
	Antibiotic TA	Myxococcus xanthus	
	Polymixin	Brevibacterium polymyxa	12
Polymeric surfactants	Alasan	Acineto bacter radioresistens	17
	Liposan	Candida lipolytica	18
	Lipomanan	Candida tropicalis	

Table-2: Examples of biological activity of biosurfactant.

Biosurfactant Type	Biological Activity	References
Surfactin	Antimicrobial and antifungal activities. Inhibition of clot formation. Haemolysis and formation of ion channels in lipid membranes. Antitumor activity against carcinoma cells.	
Rhamnolipid	Anti-adhesive activity against several bacteria and yeast strains. Antimicrobial activity against Mycobacterium tuberculosis.	40,41
Iturin	Antimicrobial activity and antifungal activity against mycosis. Enhance the electrical conductance of biomolecular lipid membranes. Destroy the membrane structure of yeast cells.	46
Pumilacidin	Antiviral activity against herpes simplex virus 1 (HSV-1).	47
Mannosylerythritol lipids	Antimicrobial, immunological and neurological properties.	48,49,50
Glycolipid	Anti-adhesive activity against several bacterial and yeast strains.	51,52
Lichenysin	Chelating properties that might explain the membrane-disrupting effect of lipopeptides.	53,54