



ANTIOXIDANT MUSHROOMS: A REVIEW

Aggarwal Preeti¹, Sharma Pushpa^{1*}, Sharma Sakshi², Aggarwal Jyoti³

¹Department of Pharmaceutical Chemistry, M. M. College Of Pharmacy, M. M. University, Mullana, Ambala, India

²Ranbaxy Laboratories, Gurgaon, India

³Rayat Institute of Pharmacy, Ropar, S.B.S Nagar, Punjab, India

Article Received on: 06/04/12 Revised on: 20/05/12 Approved for publication: 11/06/12

*Email: pushpamullana@rediffmail.com

ABSTRACT

The antioxidant properties of wild mushrooms have been extensively studied and many antioxidant compounds such as phenolic compounds, tocopherols, ascorbic acid, and carotenoids identified. The various antioxidant mechanisms of the mushroom species extracts may be attributed to strong hydrogen-donating ability, metal-chelating ability, and their effectiveness as good scavengers of superoxide and free radicals. This indicates the potential of mushrooms as panacea for many diseases and also reveals a novel potential to fight against tumors in man.

Keywords: Mushrooms, Antioxidants, Free radicals, Phenolic compounds, Tocopherols, Carotenoids.

INTRODUCTION

Mushrooms have continued to generate a lot of interest particularly in its consumption as food¹, in the cure of diseases², in bioremediation and as important items of commerce all over the world that stems from their nutritional, antioxidant and therapeutic values³⁻⁶. Mushrooms may prove to be one of the useful candidates in the search for bioactive compounds with radical scavenging activity as the fruiting body can be produced in much less time, the mycelium may also be rapidly produced in liquid culture and the culture medium can be manipulated to produce optimal quantities of active products⁷⁻¹⁰. The antioxidant properties of wild mushrooms have been extensively studied and many antioxidant compounds extracted from these sources were identified such as phenolic compounds, tocopherols, ascorbic acid, and carotenoids.

Naturally **phenolic compounds** are produced as accumulated end products from the shikimate and acetate pathways and can range from relatively simple molecules (phenolic acids, phenylpropanoids, flavonoids) to highly polymerised compounds (lignins, melanins, tannins), with flavonoids representing the most common and widely distributed subgroup, main phenolic compounds found in mushrooms were phenolic acids. Total phenols inhibit occurrence of atherosclerosis and cancer^{11,12}.

Vitamin C, a necessary nutrient, is thought to exert a protective role against various oxidative stress-related diseases such as heart disease, stroke, cancer, several neurodegenerative diseases and cataractogenesis¹³. Among simplest vitamins, it is one found in mushrooms that has been determined using HPLC coupled to UV or fluorescence detector, or following the spectrophotometer procedure based on the reaction with 2,6-dichlorophenolindophenol¹⁴.

Carotenoids, exhibiting provitamin and antioxidant roles, particularly β - carotene were found in several mushroom species. As they can undergo isomerization, they are found in trans and cis isomers in which trans isomers are more common in food and are stable. Carotenoids reactivity

depends on the length of the chain of conjugated double bonds and the characteristics of the end groups¹⁵⁻¹⁷.

L-ergothioneine is a biologically active and stable antioxidant produced by certain fungal species and mycobacterium, not in plants. The precursors to the synthesis of L-ergothioneine are the amino acids- histidine, cysteine, and methionine. Supplementation with L-ergothioneine has been shown to have a protective effect on the organs of rats against lipid peroxidation and to conserve the consumption of endogenous glutathione and α -tocopherol. The ergothioneine content of mushrooms has been reported to be in the range of 0.4-2.0mg/g (dry weight)¹⁸.

Methods for estimating Antioxidant activity

There are several methods to measure the efficiency of dietary antioxidants either as pure compounds or in food matrices. These methods focus on the different mechanisms of the antioxidant defense system, such as: the scavenging of oxygen and hydroxyl radicals, the reduction of lipid peroxy radicals, the inhibition of lipid peroxidation, or the chelation of metal ions. The interaction between free radicals (such as superoxide and hydroxyl radicals) and antioxidants can show direct evidence for antioxidants to scavenge free radicals and has been widely used to evaluate the radical scavenging ability of antioxidants¹⁹. These include: Superoxide anion radical scavenging activity; Nitric oxide scavenging activity; Thiobarbituric reactive substances (TBARS assay); 1, 3-diethyl-2-thiobarbituric acid (DETBA) method; Reducing power; 2, 2-Diphenyl-1-picrylhydrazyl (DPPH) scavenging activity; Cupric ion reducing antioxidant capacity (CUPRAC); Hydroxyl radicals scavenging activity; Chelating effects on ferrous ions; Linoleic acid assay; β -carotene bleaching inhibition; Hemolysis inhibition; SOD activity; Xanthine oxidase inhibition; Chorioallantoic membrane (CAM) assay ; Conjugated diene method; 2, 7-dichloro fluorescein diacetate (DCF) / 2, 2'-azobis(2-amidinopropane) dihydrochloride (AAPH) assay; 2, 2'-azinobis-(3-ethylbenzthiazoline-6-sulphonic acid (ABTS) radical cation scavenging activity; Hydrogen peroxide scavenging activity; Thiocyanate method; Chelating effects on cupric ions; Lipid Peroxidation method; ferric reducing

antioxidant power assay (FRAP); Trolox equivalent antioxidant capacity (TEAC)^{20,21}.

Various studies done to determine Antioxidant potential of different species of mushrooms

Determination of antioxidant activity of ten species of wild Nigerian mushrooms including *Cantharelle cibarius*, *Laccaria amethysta*, *Clitocybe odora*, *Lepista nuda*, *Macrolepiotata procera*, *Lepista saeva*, *Lactarius deliciosus*, *Laccaria laccata*, *Pleurotus ostreatus* and *Hericium erinaceus* by studying inhibition of lipid peroxidation (antioxidant activity) revealed that all ten species of wild Nigerian mushrooms have antioxidant activity as they were able to inhibit peroxidation induced in liver homogenate. *C. cibarius*, *L. amethysta*, *C. odora* and *L. nuda* species elicited higher antioxidant activity compared to the standard ascorbic acid while the remaining species inhibited peroxidation to a lower extent^{22,23}.

Recent investigations carried out showed that medicinal mushrooms occurring in South India namely *Ganoderma lucidum*, *Phellinus rimosus*, *Pleurotus florida* and *Pleurotus pulmonaris* possessed profound antioxidant and antitumor activities and also showed significant antimutagenic and anticarcinogenic activities. The ethyl acetate, methanol and aqueous extracts were effective to scavenge O²⁻ generated from the photoillumination of riboflavin, OH generated from Fenton's reaction, nitric oxide radical released from aqueous solution of sodium nitroprusside in a dose dependent manner. Methanol extract effectively reduced ferric ion in FRAP assay and scavenged DPPH radicals^{24,25}.

The potential antioxidant and reducing properties and content of antioxidant compounds, phenolics and flavonoids, of three edible mushroom species *Lentinula edodes*, *Hericium erinaceus* and *Agrocybe aegerita* from Istra region and scavenging capacity on DPPH radicals were determined. The highest extraction yield was achieved in *L. edodes* extraction and highest total phenolics and total flavonoids content, as well as TF/TP ratio were determined for *A. aegerita* dry extract and all mushroom dry extracts possessed reductive capabilities. The results indicated that the antioxidant activity of mushroom extracts highly depends on extract concentration, i.e. concentration of active compounds^{26,27}.

Antioxidative potency of commercially available mushrooms in Taiwan was studied using the DPPH method and oil-in-water emulsion oxidation. The antioxidative activities of ethanol extracts of various mushrooms in an emulsified corn oil (o/w) system at 60° C were compared. The addition of test compounds in corn oil emulsions significantly extended the induction period of lipid oxidation. Mushroom (*Agaricus bisporus*) contained significant amounts of phenolic amino acids (tyrosine, L-glutaminy-4-hydroxybenzene, 3, 4-dihydroxyphenylalanine and L-glutaminy-3, 4-dihydroxybenzene) and small amounts of vitamin C which may be responsible for the relatively high antioxidative activity. The effect of mushroom extracts on retarding emulsion oxidation was *Agaricus bisporus* > *Hypsizigus marmoreus* > *Volvariella volvacea* > *Flammulina velutipes*

> *Pleurotus eryngii* > *Pleurotus ostreatus* > *Hericium erinaceus* > *Lentinula edodes*. These results indicated that the ethanol extracts of *Agaricus bisporus* and *Hypsizigus marmoreus* are highly effective in retarding the oxidation of corn oil emulsion during storage^{28,29}.

Both the sulfation of acid *Auricularia auricular* polysaccharides (SAAAP) and the sulfation of neutral *Auricularia auricular* polysaccharides (SNAAP) derivatives possessed more powerful antioxidant competence than that of the native non-sulfated polysaccharides (AAP and NAAP) while AAP and NAAP exhibited stronger activity on scavenging both the hydroxyl radical and lipid peroxidation^{30,31}.

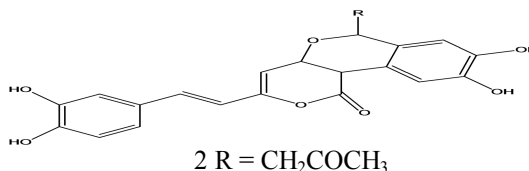
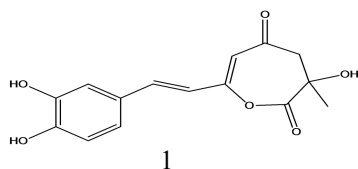
The antimicrobial activity of 90% ethyl acetate and antioxidant activity of fruiting bodies of mushrooms of four edible mushrooms *Agaricus bisporus*, *Pleurotus ostreatus*, *Volvariella volvacea* and *Pleurotus sajor-caju* was investigated. The antioxidant activity, peroxidase, number of ascorbate oxidase units and catalase activity were found to be more significant with *Agaricus bisporus* than the other edible mushrooms used in the study and no peroxidase activity was observed in *P. ostreatus*³².

The antioxidant activity of *Agaricus bisporus* was analysed by DPPH and hydroxyl (OH) radical scavenging assays that showed more scavenging activity on DPPH than OH. The phenolics composition of *A. bisporus* methanolic extracts was analysed by HPLC and found to contain rutin, gallic acid, caffeic acid and catechin which contributed to the antimicrobial and antioxidant activity. Total phenol and ascorbic acid content contributed to its antioxidant activity. This study revealed that *A. bisporus* is a natural source of antioxidant and antimicrobial agents against the tested organisms and had a potential as anticancer³³.

Ethanol extracts of edible mushroom *Agaricus bisporus* before and after boiling for antioxidant, anticancer and antimicrobial activities were investigated and found that ABTS and DPPH free radical scavenging activities were similar in both the extracts, the extracts inhibited cell proliferation of HL-60 leukemia by the induction of apoptosis and antibacterial activity against both gram positive and gram negative bacteria, as well as anticandidal activity against *Candida albicans*³⁴.

The antioxidant activity of *Agaricus brasiliensis* strains extracted with methanol as solvent on different basidiocarp maturation phases was evaluated and also the best harvest period to obtain highest antioxidant activity was determined. Strains with closed basidiocarp had higher antioxidant activity than with opened basidiocarps³⁵.

Two natural antioxidants, named inonotusin A (1) and B (2), were isolated from the methanolic extract of the fruit bodies of *Inonotus hispidus*, together with (*E*)-4-(3,4-dihydroxyphenyl)but-3-en-2-one (3), hispidin (4) and 3,4-dihydroxybenzaldehyde (5). Their structures were identified by means of extensive NMR and MS data analysis. Compounds 1, 2 and 4 exhibited significant scavenging activity against the ABTS radical cation³⁶.



followed the order crude > boiled > ethanolic extracts in hydroxyl radical scavenging activity, crude > ethanolic > boiled in DPPH radical scavenging activity, and boiled > crude > ethanolic in case of inhibition of lipid peroxidation respectively. Crude, boiled and ethanolic extracts also increased significantly nitric oxide production over control. The ethanolic extract was the most effective in relation to antioxidant activity and NOS activation property⁴⁴. Antioxidant properties of hot water and ethanolic extracts prepared from cap and stipe of *C. comatus* fruit bodies were studied. Ethanolic extract from stipe showed high antioxidant activity while extracts from cap showed better scavenging ability on DPPH than stipe ones. Ethanolic extracts were more effective in scavenging ability on hydroxyl radicals but moderate on superoxide radicals than hot water extracts. Naturally occurring antioxidant components including total phenols, tocopherols, flavonoids and polysaccharides were found in the extracts. Overall, extracts from cap were more effective for the antioxidant properties assayed⁴⁵.

The in-vitro antioxidant and ACE inhibitory activities of selected culinary-medicinal mushrooms extracted by boiling in water for 30 mins were studied. Antioxidant capacity was measured using the following assays – DPPH free radical scavenging activity, β -carotene bleaching, inhibition of lipid peroxidation, reducing power ability and CUPRAC. Antioxidant potential of each mushroom species was calculated based on the average percentages relative to quercetin. *Ganoderma lucidum* (30.1%), *Schizophyllum commune* (27.6%) and *Hericium erinaceus* (17.7%) showed relatively high Antioxidant index (AI)^{46,47}.

For the first time it was reported that *Gastrodia elata* accumulated ergothioneine (ERG) whose levels were correlated with the concentrations of ERG in *Armillaria mellea*, one of the symbiotic fungi on which the lifecycle of *Gastrodia elata*, an achlorophyllous orchid plant, is completely dependent. The contents of ERG in *G. elata* were significantly higher in actively developing tissues, such as seed capsules and newly growing corms, than in mature rhizomes. The ERG levels in rhizomes were significantly correlated with antioxidant capacities⁴⁸.

Antioxidant properties and antioxidant compounds of various extracts from the Edible Basidiomycete *Grifola Frondosa* (Maitake) currently available in Taiwan were evaluated. Ethanolic, cold-water and hot-water extracts were prepared and their antioxidant properties were investigated. Cold-water extracts showed high reducing power and chelating abilities on ferrous ions than ethanolic and hot-water extracts. For the scavenging ability on DPPH radical, *G. frondosa* extracts were effective in the following order: ethanolic > hot-water > cold-water while the hot-water extract showed high scavenging ability on superoxide anions. Total phenols, flavonoids, ascorbic acid and α -tocopherol are the major antioxidant components found due to which they display potent antioxidative properties⁴⁹.

Antioxidant activity of the ethyl acetate soluble (PdEs) and insoluble (PdEi) fractions of methanol extract of plant-parasitic macrofungus *Phellinus durissimus* (Lloyd) Roy was investigated by in vitro chemical assays and established that *P. durissimus* has significant antioxidant potential which is comparable to or better than other species of Hymenochaetales⁵⁰.

The methanolic extracts of dried *Agaricus bisporus*, *Polyporus squamosus*, *Pleurotus ostreatus*, *Lepista nuda*,

Russula delica, *Boletus badius*, and *Verpa conica* were investigated for antioxidant activity in different systems including reducing power, free radical scavenging, superoxide anion radical scavenging, total antioxidant activity, and metal chelating activities and compared to standard antioxidants BHA, BHT, and α -tocopherol. *Russula delica* showed highest percentage inhibition on peroxidation in linoleic acid system, reducing power and O²⁻ scavenging. Total phenolic compounds, α -tocopherol, and β -carotene were also determined⁵¹.

Methanol and water crude extracts from Shiitake mushroom (*Lentinus edodes*) and straw mushroom (*Volvariella volvacea*) were investigated for their antioxidant capacity in three different assays, namely, the β -carotene and linoleic acid system, DPPH radical scavenging activity, and inhibition of hemolysis of rat erythrocyte induced by peroxyl radicals. The water extract from *L. edodes* showed the most potent radical scavenging activity. Total phenolics were higher in the water extracts⁵².

ROS-generating activity in human cells and DPPH-TEAC antioxidant activity in hot water extracts of 2 groups of medicinal mushrooms from the genera *Agaricus*, *Antrodia*, *Auricularia*, *Coprinus*, *Cordyceps*, *Hericium*, *Grifola*, *Ganoderma*, *Lentinus*, *Phellinus*, and *Trametes* was determined⁵³.

The detailed mechanism whereby *G. lucidum* stimulates the catalase activity and expression was elucidated and found that methyl linoleate that is produced by *G. lucidum* stimulates the catalase expression at the transcription level⁵⁴.

An ergothioneine derivative, β -hydroxyergothioneine has been isolated from the mushroom *Lyophyllum connatum*. Ergothioneine, N-hydroxy-N', N'-dimethylurea, and connatin (N-hydroxy-N', N'-dimethylcitrulline) were also isolated. All the compounds displayed the ability to scavenge free radicals, based on DPPH radical scavenging assay. The radical scavenging activity of β -hydroxyergothioneine was very similar to that of ergothioneine. β -Hydroxyergothioneine showed the greatest protective activity against carbon tetrachloride-induced injury in primary culture hepatocytes⁵⁵. The antioxidant capacity and total phenolic content of *Agaricus brasiliensis* in two stages of maturity, young (YB) and mature (MB), have been evaluated with minor differences in the composition of phenolic compounds being detected, but with similar antioxidant activities, except for the chelating ability for ferrous ions, which was higher in MB than in YB⁵⁶.

Antioxidant properties of methanolic extracts of three species of medicinal mushrooms in Taiwan- *Ganoderma lucidum* (Ling-chih), *Ganoderma tsugae* (Sung-shan-ling-chih), and *Coriolus versicolor* (Yun-chih) were studied. Results showed that *G. lucidum* and *G. tsugae* were higher in antioxidant activity, reducing power, scavenging and chelating abilities, which was attributed to their total phenolic content⁵⁷.

Membrane ultrafiltration and subfractionation of methanol and water extracts, of two edible mushrooms *Lentinus edodes* and *Volvariella volvacea* showed that the dichloromethane subfraction of the methanol extract of *V. volvacea* and the low molecular weight (LMW) subfraction of the water extract of *L. edodes* had the highest antioxidant activity against lipid peroxidation of rat brain homogenate correlated with the phenolic content in different subfractions⁵⁸.

A new potent antioxidant vialinin A together with a known compound ganbajunin B and a mixture of ganbajunins D and

E were isolated from dry fruiting bodies of *Thelephora* vial and structure was elucidated by spectroscopic and chemical methods. This compound had strong DPPH free radical scavenging activity nearly equal to that of BHT⁵⁹.

Analysis for proximate composition, total phenols and antioxidant activity of methanolic extracts of three wild edible mushrooms (*Agaricus sp.*, *Boletus sp.*, *Macrolepiota sp.*) from North of Mexico and two commercial sp. (*Agaricus bisporus* white strain and brown strain) showed that wild mushrooms had higher phenolic content and antioxidant capacity than the commercial species⁶⁰.

CONCLUSION

Mushrooms can serve as a dietary supplement for proteins, vitamins, minerals as well as a cheap and easily accessible source for natural antioxidants for both man and/or livestock. The various antioxidant mechanisms of the mushroom species extract may be attributed to strong hydrogen-donating ability, a metal-chelating ability, and their effectiveness as good scavengers of superoxide and free radicals. This indicates the potential of mushrooms as panacea for many diseases and also reveals a novel potential to fight against tumors in man. However, more intensive and extensive investigations are needed to exploit their valuable therapeutic potential.

REFERENCES

- Chang ST. Mushroom Production in South East Asia. Mushroom Newsle Trop 1980, 4: 5-10.
- Mattila P, Suonpaa K, Piironen V. Content of vitamins, mineral elements and some phenolic compounds in cultivated mushrooms. J Agri & Food Chem 2001, 49: 2343-2348.
- Aletor VA. Compositional Studies on edible tropical specials of mushrooms. Food Chem 1995, 54: 256-268.
- Fasidi IO. Studies on *Volvariella esculenta* mass singer, Cultivation on agricultural wastes and proximate composition of stored mushrooms. Food Chem 1996, 55: 161-163.
- Okwulehie IC, Odunze ET. Evaluation of the myco-chemical and mineral composition of some tropical edible mushroom. J Sustainable Agri & Env 2004, 6(1): 63-70.
- Bano ZS. Rajarathna studies on the cultivation of pleurotus species. Mushroom J 1981, 101: 243-245.
- Chang R. Functional properties of edible mushrooms. Nutr Rev 1996, 54: 91-93.
- Chang S. Global impact of edible and medicinal mushrooms on human welfare in the 21st century: Nongreen revolution. Int J Med Mushrooms 1999, 1: 1-7.
- Lindequist U, Niedermeyer THJ, Julich WD. The pharmacological potential of mushrooms. eCAM 2005, 2: 285-299.
- Wasser SP. Medicinal properties of substances occurring in higher basidiomycete mushrooms: current perspective (review). Int J Med Mushrooms 1999, 1: 31-62.
- Apak R, Guclu K, Demirata B et al. Comparative evaluation of various total antioxidant capacity assays applied to phenolic compounds with the CUPRAC assay. Molecules 2007, 12: 1496-1547.
- Bravo L. Polyphenols: chemistry, dietary sources, metabolism, and nutritional significance. Nutr. Rev 1998, 56: 317-333.
- Halliwell B. Antioxidants in human health and disease. Annu Rev Nutr 1996, 16: 33-50.
- Klein BP, Perry AK. Ascorbic-acid and vitamin-A activity in selected vegetables from different geographical areas of the United-States. J Food Sci 1982, 47: 941-945.
- Liu RH. Potential synergy of phytochemicals in cancer prevention: mechanism of action. J Nutr 2004, 134: 3479-3485.
- Rao AV, Rao LG. Carotenoids and human health. Pharmacol Res 2007, 55: 207-216.
- Paiva SA, Russell RM. Beta-carotene and other carotenoids as antioxidants. J Am Coll Nutr 1999, 18: 426-433.
- Deiana M et al. L-ergothioneine modulates oxidative damage in the kidney and liver of rats in vivo: studies upon the profile of polyunsaturated fatty acids. Clin Nutr 2004, 23: 183-193.
- Lu JM, Peter HL, Yao QZ, Chen CY. Chemical and molecular mechanisms of antioxidants: Experimental approaches and model systems. J Cell Mol Med 2010, 14: 840-860.
- Halliwell B, Aeschbach R, Loliger J, Aruoma I. The characterization of antioxidants. Food Chem Toxicol 1995, 33: 601-617.
- Halliwell B, Whiteman M. Measuring reactive species and oxidative damage *in vivo* and in cell culture: How should you do it and what do the results mean? J Pharmacol 2004, 142: 231-255.
- Egwim EC, Elem RC, Egwuiche RU. Proximate composition, phytochemical screening and antioxidant activity of ten selected wild edible Nigerian mushrooms. Am. J. Food Nutr 2011, 1(2): 89-94.
- Adejumo TO, Awosanya OB. Proximate and mineral composition of four edible mushroom species from South Western Nigeria. African J Biotech 2005, 4(10): 1084-1088.
- Thekkuttuparambil A, Ajith, Kainoor K, Janardhanan. Indian Medicinal Mushrooms as a Source of antioxidant and antitumor agents. J Clin Biochem Nutr 2007, 40: 157-162.
- Wasser SP. Medicinal mushrooms as a source of antitumor and immunomodulatory polysaccharides. Appl Microbiol Biotechnol 2002, 60: 258-274.
- Mujic Z, Zekovic Z, Lepojevic S, Vidovic. Antioxidant properties of selected edible mushroom species. J Central European Agriculture 2010, 11: 387-392.
- Kahkonen MP, Hopia AI, Vuorela HJ. Antioxidant activity of plant extracts containing phenolic compounds. J Agricultural Food Chem 1999, 47: 3954-3962.
- Hui-yin fu, Den-en shieh. Antioxidant and free radical scavenging activities of edible mushrooms. J Food Lipids 2002, 9: 35-46.
- Bobek P, Ozdin, Kuniak L. Effect of oyster mushroom and isolated beta-glucan on lipid peroxidation and on the activities of antioxidative enzymes in rats fed the cholesterol diet. Nutr Biochem 1997, 8: 469-471.
- Hua Zhang, Zhen-Yu Wang, Lin Yang. In Vitro Antioxidant Activities of Sulfated Derivatives of Polysaccharides Extracted from *Auricularia auricular*. Int J Mol Sci 2011, 12: 3288-3302.
- Yan, P.S.; Luo, X.C.; Zhou, Q. RAPD molecular differentiation of the cultivated strains of the jelly mushrooms, *Auricularia auricula* and *A. polytricha*. World J Microbiol. Biotechnol 2004, 20: 795-799.
- Surekha Ch, Kaladhar DSVGK, Raju Srikakarlapudi JR, Haseena. Evaluation of antioxidant and antimicrobial potentiality of some edible mushrooms. Int J Adv Biotech & Res 2011, 2(1): 130-134.
- S.E. Abah and G. Abah, Antimicrobial and antioxidant potentials of *Agaricus bisporus*. Adv Bio Res 2010, 4(5): 277-282.
- Loganathan K, Jagadish V, Venkatakrishnan R, Shembhagaraman, Kaviyaranan V. Comparative study on the antioxidant, anticancer and antimicrobial property of *Agaricus bisporus* (J. E.Lange) Imbach before and after boiling. African J Biotech 2009, 8(4): 654-661.
- Francielli Mourao, Suzana Harue Umeo, Orlando Seiko Takemura, Giani Andrea Linde, Nelson Barros Colauto. Antioxidant activity of *agaricus brasiliensis* basidiocarps on different maturation phases. Brazilian J Microbio 2011, 42:197-202.
- Li-feng Zan, Jian-chun Qin, Ya-mei Zhang, Yan-hua Yao, Hai-ying Bao, Xiang LI. Antioxidant hispidin derivatives from medicinal mushroom *Inonotus hispidus*. Chem Pharm Bull 2011, 59(6): 770-772.
- Wei-Min Yang et al. Antioxidant properties of natural *p*-Terphenyl derivatives from the mushroom *Thelephora ganbajun*. Verlag der Zeitschrift fur Naturforschung 2004, 59: 359-362.
- In-Kyoung Lee et al. New antioxidant polyphenols from the medicinal mushroom *Inonotus obliquus*. Bioorg & Med Chem Letters 2007, 17: 6678-6681.
- Ricardo J, Lavitschka, Carlos R, Douglas Mascara, Priscila A et al. In vitro cytotoxicity and antioxidant activity of *Agaricus subrufescens* extracts. African J Biotech 2007, 6(9): 1144-1150.
- Afacan Makasci, Mammadov R, Dusen O, Isik HI. Antimicrobial and antioxidant activities of medicinal plant species *Ornithogalum alpigenum* stapf. from Turkey. J Med Plants Res 2010, 4(16): 1637-1642.
- Aziz Turkoglu et al. Antioxidant and antimicrobial activities of *Laetiporus sulphureus* (Bull.) Murrill. Food Chemistry 2007, 101(1): 267-273.
- Aziz Turkoglu et al. Antioxidant and antimicrobial activity of *Russula delicata* Fr: An edible wild mushroom. Eurasian J Ana Chem 2007, 2(1): 54-67.
- Mira Popovic et al. Anti-Oxidative Activity of an aqueous suspension of commercial preparation of the mushroom *Coprinus comatus* molecules 2010, 15: 4564-4571.
- Rai M, Biswas G, Chatterjee S, Subhash C, et al. Evaluation of antioxidant and nitric oxide synthase activation properties of *Armillaria Mellea* quel. J Bio Sci 2009, 1(1): 39-45.
- Bo Li, Fei Lu, Xiaomin Suo, Haijuan Nan, Bin Li. Antioxidant Properties of Cap and Stipe from *Coprinus comatus*. Molecules 2010, 15: 1473-1486.

46. Vidovic SS, Mujic IO, Zekovic ZP, Lepojevic ZD, et al. Antioxidant properties of selected *Boletus* mushrooms. Food Biophysics 2010, 5: 49-58.
47. Stamets P. Notes on nutritional properties of culinary-medicinal mushrooms. Int J Med Mushrooms 2005, 7: 103-110.
48. Eung-Jun Park, Wi Young Lee, Seung Taek Kim, et al. Ergothioneine accumulation in a medicinal plant *Gastrodia elata*. J Med Plants Res 2010, 4(12): 1141-1147.
49. Jan-Ying Yeh, Li-Hui Hsieh, Kaun-Tzer Wu and Cheng-Fang Tsai. Antioxidant Properties and Antioxidant Compounds of Various Extracts from the Edible Basidiomycete *Grifola Frondosa* (Maitake). Molecules 2011, 16: 3197-3211.
50. Suman K, Lahiri et al. Evaluation of Antioxidant Activity of Plant-parasitic macrofungus: *Phellinus durissimus* (Lloyd) Roy. Eurasian J Ana Chem 2010, 5(1): 32-45.
51. Elmastasa M, Isildaka O, Turkekulb I, Temur N. Determination of antioxidant activity and antioxidant compounds in wild edible mushrooms. J Food Composition & Analysis 2007, 20: 337-345.
52. Cheung LM, Peter CK, Cheung, Vincent EC, et al. Antioxidant activity and total phenolics of edible mushroom extracts. Food Chem 2003, 81: 249-255.
53. Song Wei, Leo JL, Griensven V. Pro- and antioxidative properties of medicinal mushroom extracts. Int J Med Mushrooms 2008, 10(4): 315-324.
54. Hyeon Yong Lee et al. Isolation and identification of an antioxidant enzyme catalase stimulatory compound from *Ganoderma lucidum*. J Biochem & Molecular Biology 2003, 36(5): 450-455.
55. Kimura C, Nukina M, Igarashi K, Sugawara Y. Beta-Hydroxyergothioneine, a new ergothioneine derivative from the mushroom *Lyophyllum connatum*, and its protective activity against carbon tetrachloride-induced injury in primary culture hepatocytes. Bio Sci Biotechnol Biochem 2005, 69(2): 357-363.
56. Francielli Mourão et al. Antioxidant activity of *Agaricus brasiliensis* basidiocarps on different maturation phases. Brazilian J Microbio 2011, 42: 197-202.
57. Mau JL, Lin HC, Chen CC. Antioxidant Properties of several medicinal mushrooms. J. Agric. Food Chem 2002, 50 (21): 6072-6077.
58. Cheung LM, Peter CK. Mushroom extracts with antioxidant activity against lipid peroxidation. Food Chem 2005, 89(3): 403-409.
59. Xie C, Koshino H, Esumi Y, Takahashi S, Kunie Yoshikawa K, et al. Antioxidant properties of several medicinal mushrooms. Bio Sci Biotechnol Biochem 2005, 69(12): 2326-2332.
60. Alvarez-Parrila, Delarosa E, Martinez LA, Gunzalez NR, Aguilav GA. Antioxidant properties of several medicinal mushrooms. Cienc Technol Aliment 2007, 5(5): 329-334.